

# Modulation of adipogenesis by *Polygonum hydropiper* L. and *P. aviculare* L. extracts

Martina Savova\*<sup>1,2</sup>, Apostol Apostolov<sup>1,2,3</sup>, Liliya Mihaylova<sup>1,2</sup>, Milen Georgiev<sup>1,2</sup>

<sup>1</sup> Department of Plant Cell Biotechnology, Center of Plant Systems Biology and Biotechnology, 4000 Plovdiv, Bulgaria

<sup>2</sup> Laboratory of Metabolomics, Institute of Microbiology, Bulgarian Academy of Sciences,  
139 Ruski Blvd, 4000 Plovdiv, Bulgaria

<sup>3</sup> Department of Plant Physiology and Molecular Biology, University of Plovdiv, 24 Tzar Assen, 4000 Plovdiv, Bulgaria

## Introduction

Obesity is among the leading health problems of the human population during the last decades. Its complex etiology obscures obesity management (Savova et al., 2021).

Nature and its plant diversity provide thousands of bioactive compounds with (un)known medicinal properties. For centuries, people have used folk remedies as single plant or plant combinations for different diseases including obesity.

*Polygonum aviculare* L. and *P. hydropiper* L. both from Polygonaceae are plants used in ethnopharmacological herbal combinations for obesity, hypertension and atherosclerosis treatment (Lee et al., 2011; Park et al., 2014).

The human Simpson-Golabi-Behmel syndrome (SGBS) preadipocyte cell strain is a relevant model for studying adipogenesis (Fischer-Posovszky et al., 2008).

The present study aimed to elucidate whether the extracts of *P. aviculare* and *P. hydropiper* areal parts affect human SGBS adipogenic differentiation and suggest their potential molecular mechanism of action.

## Materials and methods

### Plant material and extraction

Aerial parts of investigated plants were collected in September 2019 from the following locations: *P. hydropiper* - Zvanichevo village, Bulgaria, latitude 42.1908 N, longitude 24.2416 E; and *P. aviculare* -

Sandanski, Bulgaria, latitude 41.5785 N; longitude 23.2869 E. Plant samples lyophilized and grounded were subjected to ultrasound-assisted extraction in 50% methanol, at room temperature for 20 minutes. The filtered extracts were concentrated on rotary evaporator, then lyophilized and stored at -20 °C prior to use.

### Cell culture and treatment

The human SGBS cells were kindly provided by Prof. Wabitsch from the Ulm University (Ulm, Germany) and were cultured according to the optimal conditions described detailly by Tews et al. (2019). From first day of differentiation and on every fourth day with change of the media, extracts were added at concentrations of 5, 10 and 25 µg/mL. Extracts effect were compared to vehicle treatment of 0.02% DMSO.

### Cell viability

The influence of plant extracts on cell viability was assessed by MTT assay on confluent SGBS preadipocytes.

### Oil red O (ORO) adipogenesis assay

Differentiated SGBS adipocytes were fixed with 10% formalin for 10 min at room temperature, then washed twice with phosphate buffered saline and stained with the fresh filtered ORO dye solution for 15 min. Photomicrographs of the stained cells were captured using an Oxion Inverso OX.2053-PLPH inverted microscope

and DC.10000-Pro CMEX camera (Euromex, The Netherlands). Estimation of the total lipid content was done by extraction of the ORO dye from the cells with isopropanol, followed by reading the absorbance at 495 nm.

#### Free glycerol assay

Concentration of the free glycerol, released in the culture media was measured with Adipolysis Assay Kit (MilliporeSigma) according to the manufacturer's instructions.

#### Quantitative real-time polymerase chain reaction (RT-qPCR)

Total RNA was isolated and reverse transcribed to cDNA. Expression of the major adipogenic genes was detected with RT-qPCR as described by Savova et al. (2021).

#### Western blot analysis

Protein isolation and Western blotting were performed as described previously (Savova et al., 2021).

## Results and discussion

Plant extracts from *P. aviculare* and *P. hydropiper* did not affect cell viability of SGBS cells.

*Polygonum aviculare* anti-atherosclerotic effect has been evaluated in vivo in ApoE knockout model on high-fat diet fed mice (Park et al., 2014). The anti-obesity effect of *P. hydropiper* has been assessed in vitro in murine 3T3-L1 adipocytes. It was suggested that the flavonoids - isoquercitrin and isorhamnetin - are responsible for activation of Wnt/ $\beta$ -catenin pathway and inhibition of adipocyte differentiation (Lee et al., 2011). Consistently, our data suggest that both plants from Polygonaceae family modulate adipogenesis.

Treatment with both *P. aviculare* and *P. hydropiper* extracts showed significant dose-dependent decrease in lipid accumulation in human SGBS adipocytes. Moreover, decrease in basal lipolysis as reduction of released glycerol in the culture media was observed. Analysis of relative mRNA expression revealed significant downregulation of *CEBPA*.

Decrease in protein abundance of PI3K was observed upon all treatments, while PPAR $\gamma$  protein expression was

decreased by both extracts in the highest concentration.

## Conclusion

*Polygonum hydropiper* and *P. aviculare* modulate adipogenesis and could be employed as natural sources of bioactive molecules or in combination with plant-derived compounds with anti-obesogenic activity.

## Acknowledgements

This project for establishment of CPSBB has received funding from the European Union's Horizon 2020 research and innovation programme, project PlantaSYST (SGA No 739582 under FPA No. 664620), and the BG05M2OP001-1.003-001-C01 project, financed by the European Regional Development Fund through the "Science and Education for Smart Growth" Operational Programme.

The SGBS cell strain was provided by courtesy of Prof. Dr. Martin Wabitsch (University of Ulm, Germany).

## References

- Fischer-Posovszky, P., Newell, F.S., Wabitsch, M., Tornqvist, H.E., 2008. Human SGBS cells - a unique tool for studies of human fat cell biology. *Obes. Facts.* 1, 184–189. DOI: <https://doi.org/10.1159/000145784>.
- Lee, S.H., Kim, B., Oh, M.J., Yoon, J., Kim, H.Y., Lee, K.J., Lee, J.D., Choi, K.-L., 2011. *Perscaria hydropiper* (L.) Spach and its flavonoid components, isoquercitrin and isorhamnetin, activate the Wnt/ $\beta$ -catenin pathway and inhibit adipocyte differentiation of 3T3-L1 cells. *Phytother. Res.* 25, 1629–1635. DOI: <https://doi.org/10.1002/ptr.3469>.
- Park, S.H., Sung, Y.Y., Nho, K.J., Kim, H.K., 2014. Anti-atherosclerotic effects of *Polygonum aviculare* L. ethanol extract in ApoE knock-out mice fed a Western diet mediated via the MAPK pathway. *J. Ethnopharmacol.* 151, 1109–1115. DOI: <https://doi.org/10.1016/j.jep.2013.12.021>
- Savova, M., Vasileva, L., Mladenova, S., Amirova, K., Ferrante, C., Orlando, G., Wabitsch, M., Georgiev, M., 2021. *Ziziphus jujuba* Mill. leaf extract restrains adipogenesis by targeting PI3K/AKT signaling pathway. *Biomed. Pharmacother.* 141, 111934. DOI: <https://doi.org/10.1016/j.biopha.2021.111934>.
- Tews, D., Pula T., Funcke, J.B., Jastroch, M., Keuper, M., Debatin, K.M., Wabitsch, M., Fischer-Posovszky, P., 2019. Elevated UCP1 levels are sufficient to improve glucose uptake in human white adipocytes. *Redox. Biol.* 26, 101286. DOI: <https://doi.org/10.1016/j.redox.2019.101286>.