

Formulation and evaluation of a solid self-emulsifying drug delivery system containing cefuroxime axetil

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Introduction

One of the biggest problems in formulation of oral pharmaceutical dosage forms is the lipophilic nature of the drug substances, therefore, various formulation strategies have been explored in recent years such as usage of lipid solutions, emulsions and emulsion concentrates prepared as physically stable formulations suitable for encapsulation of poorly soluble drugs (Rodriguez-Aller et al., 2015).

Formulation approaches, such as self-emulsifying drug delivery systems (SEDDS) have been recently used to overcome drug solubility problems (Vasconcelos et al., 2017) by using isotropic multi component system composed of oil, surfactant and co-surfactant/co-solvent, which form micro- or nano-emulsion in the presence of water.

Soft gelatine capsules are simple and commonly used dosage forms for encapsulation of liquid SEDDS, nevertheless, this technology have some limitations *i.e.* manufacturing and packaging process difficulties, stability of the final drug product, etc. Due to these reasons, attention has been focused on formulation of solid SEDDS in form of tablets (uncoated or film-coated tablets) (Joyce et al., 2018). In that direction, various formulation strategies have been used for solidification of SEDDS, among which the simplest approach is adsorption of the SEDDS to solid carriers–excipients, commonly used in tablet formulation (Mandić et al., 2017). The purpose of the study was formulation of a solid

SEDDS containing the poorly soluble cefuroxime axetil in a form of a conventional tablet.

Materials and methods

Cefuroxime axetil (CA) was supplied from Orhid Chem. Pharm. Ltd. (India). Olive oil, talc (Pardeck[®] Lub Talk Emprove), methanol and citric acid monohydrate were obtained from Merck KgaA, Germany. Microcrystalline cellulose (MCC-Avicel[®] PH 112) and croscarmellose sodium (CS-AcDiSol[®]) were supplied from FMC Biopolymer, USA. Polyvinylpyrrolidone (PVP-Kollidon[®] 30F) and Polyvinylpyrrolidone, crosslinked (PVP-C-Kollidon[®] CL) were supplied from BTC Chem. Distrib., Germany. Polysorbate 80 (Tween 80) was obtained from Croda Europe Lim., France, polyethylene glycol (PEG 400) from Clariant, Germany, colloidal anhydrous silica (CAS-Aerosil[®] 200) from Pharma Evonik Ind., Germany, magnesium stearate (MS) from FaciSpA-Carasco GE, Italy and lactose monohydrate (LM-Tablettose[®] 100) from Meggle, Germany.

Preparation of solid SEDDS granules and tablets

CA was successfully incorporated in the mixture of olive oil, PEG 400, Tween 80 and citric acid monohydrate (data are not presented). Incorporation of the SEDDS onto solid drug carries was done using a high shear granulation technique (all samples

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contained SEDDS 57% and CAS 18%; additionally, granules were prepared with: *S1*: MCC 13%, PVP 2.2%, PVP-CL 9%; *S2*: MCC 13%, PVP 2.2%, CS 9%; *S3*: LM 13%, PVP 2.2%, CS 9% and *S4*: MCC 13%, PVP-CL 9.5%; impeller speed 400 rpm, chopper speed 1000 rpm, 5 min; Diosna P1/6, Diosna Dierks & Söhne GmbH, Germany). Obtained granules (*S1-4*) were sieved (mesh size 18) and blended with the lubricant and glidant (Talc 0.5%, MS 0.5%; 25 rpm, 3 min; Erweka PM5, Erweka GmbH, Germany). Flow properties of the final blends were evaluated according to Ph. Eur. 9.0 (2.9.36 and 2.2.32, respectively). Prepared granules were compressed into oblong, biconvex, white tablets with length of 21 mm on rotary tablet press (Korsch Pro XL100, Germany). The tablets were characterized for: mass and uniformity of mass (Ph. Eur. 9.0 (2.9.5); Sartorius Secura 224-1 CEU, Sartorius AG, Germany), hardness, diameter and thickness (Erweka Hardness Tester 425 TD, Erweka GmbH, Germany), friability (Ph. Eur. 9.0 (2.9.7); Erweka TAR 100, Erweka GmbH, Germany) and disintegration (Ph. Eur. 9.0 (2.9.1); Erweka ZT322, Erweka GmbH, Germany). Assay analyses of CA were done using the HPLC systems (Agilent 1200 Binary Series, DAD detector, Germany and Hitachi Elite[®] Lachrom, Hitachi, USA), column (Inertsil[®] ODS-2, 4.6x150 mm, 5 μ m, GL Science, Japan). As a mobile phase, 20% methanol and 80% water purified were used, with chromatographic conditions: mobile phase flow of 1 mL/min, column temperature of 40 °C, injection volume of 100 μ L at 281 nm. *In vitro* CA release from the tablets ($n = 3$) was carried out with USP Apparatus II using 900 mL, 0.07 N HCl as dissolution media (Varian Vankel 7025 Model 115/230, Varian, USA) at 37 \pm 0.5 °C and 100 rpm. At predetermined time intervals (after 5, 10, 15, 20, 30 and 45 min.), 10 mL were withdrawn and filtrated (0.20 μ m) and were analyzed by above mentioned HPLC method.

Results and discussion

Prepared granules showed fair (*S1-2*) to good flow (*S3-4*) properties, considering the high oil loading in the formulations. Characterization of the physical properties of the tablets showed that all obtained results are within the predetermined acceptance criteria (tablets mass of 1100 mg \pm 5%

with 21 \pm 0.3 and 6.5 \pm 0.2 mm for tablet diameter and thickness, respectively, 2-5 kP hardness, friability of max 1% and disintegration time of max. 15 min.

Determined CA content and the drug release studies from different tablets as solid SEDDS formulations showed that sample *S3* released ~ 87% of CA during the period of 15 min. and more that 96% after 45 min. (99.6% CA content), thus complying with the USP specification for conventional CA tablets. In comparison, the *in vitro* release of CA (125 mg) as a powder was 8 and 13%, respectively, in the same time intervals and under the same test conditions.

Conclusion

Conventional immediate release tablets containing SEDDS of CA were prepared using simple solidification with adsorption of the SEDDS onto solid carrier containing different excipients. Based on *in vitro* dissolution studies, selected sample will be further evaluated for formulation optimization.

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